SPECIAL REPORT

Top Priorities for Cerebroprotective Studies

A Paradigm Shift

Patrick Lyden[®], MD; Alastair Buchan, MD; Johannes Boltze[®], MD, PhD; Marc Fisher, MD; on behalf of the STAIR XI Consortium*

ABSTRACT: Despite years of basic research and pioneering clinical work, ischemic stroke remains a major public health concern. Prior STAIR (Stroke Treatment Academic Industry Roundtable) conferences identified both failures of clinical trial design and failures in preclinical assessment in developing putative ischemic stroke treatments. At STAIR XI, participants in workshop no. 1 Top Priorities for Neuroprotection sought to redefine the neuroprotection paradigm and given the paucity of evidence underlying preclinical assessment, offer consensus-based recommendations. STAIR proposes the term brain cytoprotection or cerebroprotection to replace the term neuroprotection when the intention of an investigation is to demonstrate that a new, candidate treatment benefits the entire brain. Although "time is still brain," tissue imaging techniques have been developed to identify patients with both predicted core injury and penumbral, salvageable brain tissue, regardless of time after stroke symptom onset. STAIR XI workshop participants called this imaging approach a tissue window to select patients for recanalization. Elements of the neurovascular unit show differential vulnerability evolving over differing time scales in different brain regions. STAIR proposes the term target window to suggest therapies that target the different elements of the neurovascular unit at different times. Based on contemporary principles of rigor and transparency, the workshop updated, revised, and enhanced the STAIR preclinical recommendations for developing new treatments in 2 phases: an exploratory qualification phase and a definitive validation phase. For new, putative treatments, investigators should carefully characterize the mechanism of action, the pharmacokinetics/pharmacodynamics, demonstrate target engagement, and confirm penetration through the blood-brain barrier. Before clinical trials, testing of candidate molecules in stroke models could proceed in a comprehensive manner using animals of both sexes and to include significant variables such as age and comorbid conditions. Comprehensive preclinical assessment might include multicenter, collaborative testing, for example, network trials. In the absence of a proven cerebroprotective agent to use as a gold standard, however, it remains speculative whether such comprehensive preclinical assessment can effectively predict clinical outcome.

Key Words: blood-brain barrier
brain
ischemic stroke
neuroprotection
reperfusion

Despite years of basic research and pioneering clinical work, ischemic stroke remains a major public health concern. Basic and clinical research has produced rational approaches for neuronal protection,¹ acute reperfusion therapies,²³ various devices for mechanical revascularization,³ and strategies for regeneration of brain tissue damaged by ischemia.^{4,5} All these innovations were based on widely accepted scientific principles and data from preclinical studies, yet no previous candidate neuroprotective therapy has successfully entered clinical practice. The reasons for this clinical-translational failure remain uncertain, but prior STAIR (Stroke Treatment Academic Industry

Roundtable) conferences identified failures of clinical trial design, as well as failures in the preclinical assessment approaches. At STAIR XI, participants in workshop no. 1 Top Priorities for Neuroprotection sought to redefine the neuroprotection paradigm. Participants aimed to revise the STAIR recommendations for preclinical assessment in light of new information about differential vulnerability in the neurovascular unit and its dynamic time course, the emergence of new, pleiotropic agents to protect brain, and renewed commitment to scientific rigor.

In most prior efforts to develop stroke therapy, an emphasis was placed on testing single-action,

The opinions expressed in this article are not necessarily those of the editors or of the American Heart Association.

^{*}A list of all STAIR XI Consortium contributors is given in the Data Supplement

This manuscript was sent to Midori A. Yenari, Guest Editor, for review by expert referees, editorial decision, and final disposition.

The Data Supplement is available with this article at https://www.ahajournals.org/doi/suppl/10.1161/STROKEAHA.121.034947.

For Sources of Funding and Disclosures, see page xxx.

^{© 2021} American Heart Association, Inc.

Stroke is available at www.ahajournals.org/journal/str

Nonstandard Abbreviations and Acronyms

APC	activated protein C	
ECASS	European Cooperative Acute Stroke Study	
GABA	gamma aminobutyric acid	
NINDS	National Institute of Neurological Disorders and Stroke	
NVU	neurovascular unit	
PAR	protease-activated receptor	
PSD	postsynaptic density	
r-tPA	recombinant tissue-type plasminogen activator	
STAIR	Stroke Treatment Academic Industry Roundtable	

single-target agents. This approach ignored the fact that ischemia produces a plethora of pathological pathways proceeding both in series and in parallel.⁶ Workshop participants propose that a multiple-action, multiple-target approach for ischemic stroke could have a higher likelihood of success in patients with stroke. Future ischemic stroke treatment might target the neurovascular unit using pleiotropic agents acting at multiple points in the ischemic cascade and at different times. For example, the novel drug, 3K3A-APC (activated protein C), acts on the PAR-1 (protease-activated receptor 1) found on endothelial cells, astrocytes, pericytes, and neurons, allowing a treatment strategy that targets the entire neurovascular unit.⁷ Alternatively, an agent might be developed to act on a key single target that serves as a hub for multiple cytodestructive downstream signaling pathways. NA-1, by acting specifically on the PSD (postsynaptic density)-95 protein, disrupts glutamate receptor interactions with multiple effector molecules potentially reducing ischemic cell injury via several downstream mechanisms.^{8,9} Both of these drugs are in clinical development to determine if significant benefit accrues with their use.^{10,11}

Despite advances in revascularization therapies, there remains a large unmet need in ischemic stroke treatment: not all patients treated with thrombolysis or mechanical thrombectomy recover full or substantial function. Also, the risk of hemorrhage after recanalization therapiesalthough uncommon-dissuades some practitioners from using them.¹² Thus, adjuvant treatments are needed to complement recanalization therapies. In past stroke treatment trials, conducted before the advent of mechanical thrombectomy as a treatment option, it is likely that many patients failed to reperfuse; the candidate adjuvant treatments were tested in the more challenging setting of permanent rather than transient brain ischemia and thus failed to show benefit in clinical trials. In modern clinical stroke trial design, candidate adjuvant therapy can be studied in concert with recanalization. In patients with large vessel occlusion, >80% treated with mechanical thrombectomy do substantially recanalize, although downstream reperfusion may be less than complete.¹³ Thus, modern clinical trials using thrombolysis for all indicated patients and mechanical thrombectomy for large vessel occlusion patients ought to have a much greater probability that the candidate drug will succeed. Bridging agents—given to slow ischemic injury progression and increase volume of salvageable tissue at the time of definitive recanalization—might now demonstrate benefit. Studies of agents intended to prevent post-reperfusion injury will now have many more eligible patients for testing.

Once an adjuvant treatment shows benefit in combination with recanalization, then, subsequent additional trials could assess for benefit in patients who are ineligible for recanalization therapies. Although there is a tremendous need for treatment targeting patients ineligible for recanalization therapies, workshop participants concluded that for the time being clinical trials should target patients undergoing recanalization to optimize the chance of demonstrating a beneficial effect.

After presentations on the above topics, workshop participants developed the following recommendations.

CHANGING THE PARADIGM: NOMENCLATURE

For decades, stroke investigators used the term neuroprotection in 2 ways. Either we meant protection of the entire brain during injury—usually ischemic stroke but also brain trauma and cerebral ischemia due to cardiac arrest—or we meant the salvage of neurons during injury in cell culture with glutamate application, other chemical injury, or simulated ischemia with oxygen-glucose deprivation. Using these in vitro neuronal protection models, we assumed that treatments that protected neurons in cell culture would translate into agents that would protect whole brain in stroke models, and subsequently translate into effective treatments for human stroke victims. This paradigm has failed to date, perhaps because the neurovascular unit encompasses more than just neurons.

Recent data confirms that the several elements of the neurovascular unit (NVU) behave differently during ischemia.¹⁴ The NVU consists of several cell elements: neurons, astrocytes, endothelial cells, pericytes, oligodendroglia, and microglia are the most commonly studied; these cells all interact with blood and peripheral immune cells. The phenomenon called selective vulnerability was described many years ago.^{15–17} Neurons were found to be most vulnerable, followed by astrocytes, followed by endothelial cells; this hierarchy of vulnerability was speculated to derive from the relative distances to each cell type from the nearby microcirculation.¹⁸ Regional differences in the excitotoxic ratio—relative densities of excitotoxic glutamate versus inhibitory gamma aminobutyric acid (GABA)–ergic synapses–also relate to selective vulnerability.¹⁵ Recently, this fundamental

concept has been updated into the context of the NVU where elements of the NVU show differential vulnerability (Figure 1).¹⁴ Unlike selective vulnerability, which relates to regional differences in cell death during stroke, differential vulnerability refers to the innate susceptibility of NVU elements to ischemia in monocellular cultures. Here, neurons are still most vulnerable, but astrocytes exhibit the greatest resistance to ischemia (Figure 1); the mechanisms underlying differential vulnerability remain obscure.

In addition to differential vulnerability, several laboratories have shown that neurons signal astrocytes for help during injury but the identity of the neuronal help-me signal remains to be determined with certainty¹⁹; candidates include thrombin²⁰ and β2-estradiol.²¹ Furthermore, after help-me signaling from neurons, astrocytes activate to protect adjacent neurons–a response now well-documented by several groups.^{22–24} Studies of the mechanism of astrocyte-neuron paracrine protection could lead to the rational design of new therapeutic agents.^{25–29}

The clinical importance of differential vulnerability/ response remains to be proven but could be quite significant. In translational stroke research, investigators have applied cerebroprotective agents as if all elements of the NVU respond similarly. This presumed uniform response across all NVU elements may be wrong, and in fact, may partly explain some prior failures of human stroke clinical trials. To illustrate, therapeutic hypothermia was tested with respect to differential vulnerability. Brief, deep therapeutic hypothermia protected neurons during oxygen-glucose deprivation and the brain during rodent middle cerebral artery occlusion, but long durations of therapeutic hypothermia inhibited astrocytes, and abrogated the protective paracrine astrocyte response, thereby increasing neuronal death.¹⁴ In other words, a cytoprotective strategy intended to protect one cell element of the NVU may cause unintended and significant harm in another element: agents that impact one element of the NVU beneficially may impact another element guite differently, causing unexpected and worse outcomes in clinical trials.

In light of the above considerations, at the prior STAIR X, the term brain cytoprotection was proposed to replace the term neuroprotection.³⁰ At STAIR XI, this term was endorsed and further refinements were made to the proposed terminology.

Recommendation 1

- Cerebroprotection should be clearly defined and stated when the intention of an investigation is to demonstrate that a new candidate treatment benefits the entire brain as measured by either tissue volume, neurologic function, or preferably both. Brain cytoprotection connotes the same intent.
- 2. The term neuroprotection should be avoided when the investigator seeks to demonstrate pan-cellular brain protection.

SPECIAL REPORT

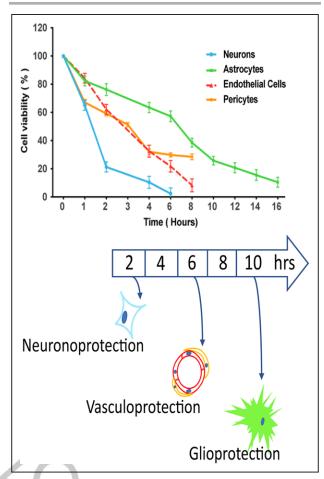


Figure 1. Differential vulnerability.

The several elements of the neurovascular unit each exhibit different susceptibility to oxygen-glucose deprivation.¹⁴ Monocellular cultures underwent varying durations of oxygen-glucose deprivation, and viability was measured 24 h later. Neurons showed greatest vulnerability, astrocytes least vulnerability, and pericytes/endothelial cells were intermediate. The term brain cerebroprotection includes protection of each element of the neurovascular unit: neuronoprotection, vasculoprotection, and glioprotection. **Upper** part: reprinted from Lyden et al¹⁴ with permission. Copyright ©2019, SAGE Publications, Ltd. **Lower** part: ©Patrick Lyden, MD.

- 3. Brain cytoprotection (Figure 1).
 - i. Neuronoprotection refers to the preservation/ protection of neurons, either in cell culture or using cell identification techniques in vivo, as in selective neuronal vulnerability.
 - II. Glioprotection refers to the preservation/protection of glia, mainly astrocytes but also oligodendroglia, in cell culture or in vivo.
 - III. Vasculoprotection refers to preserving bloodbrain barrier and reducing vascular leakage in vivo or the preservation of endothelial cells and pericytes in cell culture.
- 4. Timing: as shown in Figure 1, each version of brain cytoprotection will proceed on different time scales. Investigators must anticipate differential response in the NVU, depending on timing, and alter administration schedules accordingly.

OPPORTUNITY WINDOWS

The workshop participants next focused on approaches to offering therapy to patients when it can be most beneficial, that is, avoiding futile or even hazardous interventions. For the few decades after publication of the National Institute of Neurological Disorders and Stroke (NINDS) r-tPA (recombinant tissue-type plasminogen activator) for Acute Stroke Trial, great attention focused on time windows because intravenous thrombolysis functioned best if used soon after stroke symptom onset (Figure 2). The mantra "time is brain" reflected the importance of urgent recanalization, but clinicians and investigators recognized that in many patients some injured brain tissue might remain salvageable after the 3-hour time window used in the original NINDS study or subsequently even after the 4.5-hour time window used in the ECASS 3 trial (European Cooperative Acute Stroke Study).^{31,32} As a result, tissue imaging techniques were developed to identify patients with both predicted core injury and penumbral, salvageable brain tissue, regardless of time after stroke symptom onset. Tracer perfusion techniques allow imaging of brain areas with slow blood

flow (potentially salvageable) or zero flow (presumably completed infarction), and this perfusion mismatch correlates generally with the ischemic penumbra.33 Even closer to direct tissue imaging, magnetic resonance imaging allows demonstration of brain areas showing diffusion restriction, correlating with ischemic injury, and brain areas showing hyperintensity on Fluid Attenuated Inversion Recovery which roughly correlates with irreversibly injured brain.34 STAIR XI workshop participants called this imaging approach a tissue window to select patients for recanalization³⁵ (Figure 2). Recent studies suggest benefit for r-tPA beyond 4.5 hours of symptom duration, or due to unwitnessed onset using these imaging-based patient selection techniques. Additional studies demonstrated benefit for endovascular thrombectomy at 6 to 24 hours, also using imaging-based patient selection.34-37

Recommendation 2

1. Elements of the NVU show differential vulnerability evolving over differing time scales in different brain regions, suggesting the term target window. In the

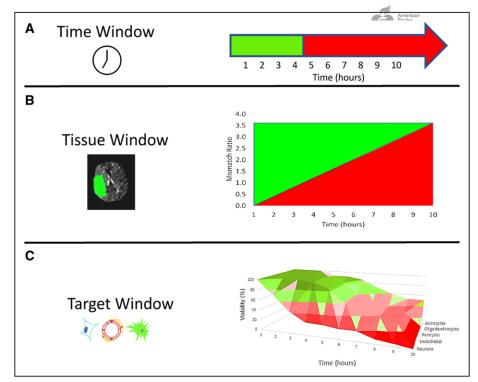


Figure 2. Opportunity windows.

Scheme for understanding the historical evolution of successful treatment for acute ischemic stroke. Green indicates beneficial outcome, red indicates death or disability, and pink indicates a transition epoch where outcome may not be best, but not worst. These graphs are provided for illustration only, are not data-derived, and are not intended to suggest specific recommendations. **A**, Time window. Initially, thrombolysis and thrombectomy (together known as recanalization therapy) could only be targeted using clock time, defined as the time since the patient was last known well and free of new stroke deficits. **B**, Tissue window. The development of perfusion imaging allowed for the estimation of perfusion mismatch and insight into the volume of salvageable tissue. Magnetic resonance-based techniques allowed even more direct imaging of tissue injury. Both perfusion and tissue injury methods allow recanalization therapy to target salvageable tissue, rather than depending on clock time. Note: Despite the appearance of the figure, the relationship between time and mismatch is likely nonlinear. **C**, Target window. In the future, both recanalization and brain cerebroprotection therapy might target different elements of the neurovascular unit at different times, knowing the differential progression of pathology in differentially vulnerable regions, or different brain regions using focal, targeted delivery (not illustrated). ©Patrick Lyden, MD.

SPECIAL REPORT

future, investigators could develop therapies that target the different elements of the NVU at different times.

OPTIMAL DESIGNS FOR PRECLINICAL EVALUATION

In the past 5 years, 2 significant developments raise new hope for investigators developing brain cerebroprotectants: the appearance of new compounds with multiple mechanisms of action and the promulgation of new standards for rigorous preclinical development.^{38–41} Workshop participants agreed to modify the existing STAIR preclinical recommendations to incorporate new developments.

In Table 1 the original STAIR guidelines⁴² (1999) are compared with the modified version^{30,43} (2009) along with extra recommendations that were added at subsequent STAIRS. In Table 2, the workshop participants updated, revised and enhanced the STAIR preclinical recommendations. It should be noted, however, that in the absence of any proven cerebroprotective stroke therapy, these recommendations cannot be ranked with respect to any supportive evidence base.

Qualification Phase

Stroke models can be used for 2 very different purposes, as codified in Table 2. For understanding a new candidate stroke therapy, that is, exploratory research,

Table 1.	STAIR Recommendations for Preclinical Stroke Treatment by Epoch
----------	---

Dose response curve	The drug should exhibit different effects over a range of doses
Time window	Determine the maximum delay to treatment after which the treatment fails.
Permanent then transient occlusion	Treatment should be studied in both permanent and transient occlusion (reperfusion) models
Blinded, physiologically controlled, reproducible studies	Physiological variables should be monitored and maintained. Laser Doppler flow drop of at least 60% should be required.
Histological and behavioral outcomes	Outcomes should include both estimates of stroke lesion volume and behav- ioral outcomes
Sex*	Consider studying treatments in both males and females.
Multiple mechanisms*	Consider combinatorial approaches
Rodent then gyrencephalic species	Demonstrate efficacy in at least 2 species
3. Additional STAIR recommendations (200)9)
Sample size calculation	Report standard deviation and predicted estimated effect size
Inclusion/exclusion criteria	For example, required drop in laser Doppler flowmetry or symptom severity
Randomization	Group allocation should be randomized
Allocation concealment	Surgeon performing stroke remains unaware of treatment assignment
Reporting on excluded animals	Account for all dropout animals
Blinded assessment of outcome	Behavioral raters, image analysts unaware of treatment assignment
Reporting of investigator or institutional conflicts of interest	Any relationship that could be perceived to introduce conflict of interest should be disclosed
C. Extra recommendations	
Age	Assess treatment effects in aging animals
Sex	Test in both sexes
Comorbidities	Test in the presence of hypertension or diabetes
Multiple laboratories	Results should be confirmed in more than one laboratory
D. STAIR XI Extra recommendations	
Mechanism	Studies of basic mechanisms should be clearly identified and differentiated from preclinical assessments for efficacy. End points should reflect the mech nism under study
Sample size	Mechanistic studies must be adequately powered for the chosen end point and predicted effect size
Preclinical assessments	Studies seeking to qualify a candidate treatment for clinical trials should be

Past STAIR Preclinical Recommendations. Original recommendations from STAIR 1 (published in 1999) were updated in 2009. Extra recommendations were added at subsequent STAIRs, including STAIR XI. STAIR indicates Stroke Treatment Academic Industry Roundtable.

*Note: Although sex and multiple mechanisms were addressed in the 1999 version, these are not traditionally considered part of the original STAIR Preclinical Recommendations.

Table 2. STAIR XI Consolidated Recommendations 2021

A. Candidate treatment qualification			
Dose response	Treatment effect varies with changes in dose		
Time window	Treatment remains effective when administered after clinically relevant delay times		
Histological and behavioral outcomes	Beneficial effects can be demonstrated using measures of behavior and tissue damage		
Target engagement	Candidate treatment reaches presumed target and causes expected physiological effects		
Barrier penetration	Candidate treatment enters brain		
B. Preclinical assessment	and validation		
Sample size	Sample size should be prespecified based on known or assumed SD and predicted effect size		
Inclusion/exclusion criteria	Effective MCA occlusion is confirmed using laser Doppler or other flowmetry or symptom severity		
Randomization	Animals are randomized before initiation of any study procedures		
Allocation concealment	Surgeon performing stroke remains unaware of treatment assignment		
Reporting on excluded animals	Subjects lost at each experimental step after randomization are summarized		
Blinded assessment of outcome	Investigators remain unaware of treatment assignment during all assessments		
Age	Consider effects of age on outcome		
Sex	Males and females should be assessed. Dose- response differences between sexes should be determined		
Comorbidities	Ideal models of stroke comorbid conditions (eg, diabetes or hypertension) need to be refined		
Multiple laboratories	Concordant effects should be demonstrated across multiple laboratories using similar methods.		
Gyrencephalic species	Demonstration of efficacy in gyrencephalic species, particularly nonhuman primates may contribute to predicting clinical efficacy		
Circadian effects	Preclinical testing of therapies during the awake phase of rodent models should be considered.		
Reporting of investiga- tor or institutional conflicts of interest	Investigator and institution conflicts are reported and managed		

Revised STAIR Recommendations. After STAIR XI all prior recommendations were revised, consolidated, and updated. Revised STAIR recommendations are separated for 2 experimental purposes. A: Candidate treatment qualification, which means early research and development of a novel, putative treatment. B: preclinical assessment and validation, which means demonstrating efficacy in stroke models that have a likelihood of predicting success in subsequent patient clinical trials. Note: STAIR recommendations are not guidelines or protocols, but rather consensus suggestions from an expert panel for investigators to consider. MCA indicates middle cerebral artery; and STAIR, Stroke Treatment Academic Industry Roundtable.

we recommend that investigators should seek to understand the mechanism of action, the pharmacokinetics/ pharmacodynamics, demonstrate target engagement, and confirm penetration through the blood-brain barrier. These activities comprise Candidate Treatment Qualification as summarized in Table 2. In the qualification phase, simple animal models may be helpful in demonstrating target engagement, blood-brain barrier transit, and other useful mechanistic studies. For reasons of expense and time, early screening studies of candidate treatments may be done in simpler stroke models, for example, younger animals, but later should proceed to more costly studies of aged animals. In these screening studies, rodents are commonly used, typically without comorbidities. Both permanent and temporary occlusion models should be investigated with histological, behavioral, and biomarker end points.

Validation Phase

Before embarking on an expensive clinical development program, investigators and regulators often seek assurance that the candidate treatment shows signals of efficacy and safety. One way to demonstrate promising signals is to use a clinically relevant animal stroke model. Prior STAIR conferences have recommended that preclinical assessment be approached with rigor and a commitment to scientific best practice, summarized in Table 2. In this phase, testing of candidate molecules in animal models could proceed in a comprehensive manner using animals of both sexes and to include significant variables such as age and comorbid conditions. Comprehensive preclinical assessment might include multicenter, collaborative testing, for example, network trials. In the absence of a proven cerebroprotective agent to use as a gold standard, however, it remains speculative whether such comprehensive preclinical assessment can effectively predict clinical outcome.

Intense analysis of previous preclinical development programs in stroke and neurodegeneration have identified key problems that should be addressed, starting with a variety of biases that have limited the generalizability and validity of animal research in general, and stroke modeling specifically.^{44,45} The workshop participants recommend that investigators commit to significant improvement and advancement of preclinical development by implementing the following technical innovations in a preclinical stroke testing network: central randomization, masking treatment assignment, power analysis and rational sample sizing, replication in multiple laboratories, study with key factors that impact outcome, for example, diabetes, hypertension, age, sex. Again it must be noted that no support is available to rank these recommendations with respect to a level of evidence.

In addition to the above innovations, simulations have suggested the superiority of multisite trials over larger single-laboratory studies.⁴⁶ The multisite approach improves the external validity and may improve the likelihood of clinical success. Such collaborative science will require bringing together multiple sites to collaborate, agree on, and implement difficult protocols. Molecules that demonstrate promise in early screening studies could be investigated more completely in well-designed and performed multisite studies before proceeding to clinical trial evaluation. Furthermore, multisite trials necessarily increase the heterogeneity of the study population, a key feature of human clinical trials usually missing in preclinical animal models.⁴⁶ That such heterogeneity improves clinical predictability remains a hypothesis to be tested.

The Stroke Preclinical Assessment Network (SPAN) was created by NINDS to develop an approach to studying putative stroke therapies in a manner that addresses the above issues. SPAN uses a novel system of distributed, masked evaluation. The SPAN platform allows each investigative lab to upload outcome data (images or behavior video recordings) in a masked fashion. Then, recordings are assigned for review to other site(s) for masked evaluation. A centralized database allows for data monitoring and quality control. The resultant data is summarized, analyzed, and when all data is locked, the code will be broken for analysis. This novel approach allows for a secure, masked, highly cost-efficient, tightly managed system with built-in central quality control.

Randomization is central to eliminating bias and establishing rigor. Prior studies suggest that simple, benchside strategies (eg, coin flipping, alternating odd/even days) retain some susceptibility to bias depending on the implementation. Successful parallel testing of multiple compounds with an adaptive strategy requires sophisticated approaches. Several authorities recommend that randomization occur before the stroke surgery (regardless of ischemia method).^{39,47} SPAN uses centralized randomization of subjects at enrollment, before stroke surgery.

Recommendation 3

- Preliminary studies of treatments for stroke should include dose/response effect, time window characterization, behavioral and histological outcomes, target engagement, and blood-brain barrier penetration (Table 2, A). We recommend that candidate treatments showing promise in such qualifying studies might then be tested more intensively (Table 2, B), including consideration of these following factors:
- 2. Sample size: Investigators should know in advance the standard deviation and predicted effect size of the treatment they plan to study. Sample size should be estimated to detect the predicted effect size with reasonable power.
- 3. Inclusion/exclusion criteria: Subjects must be excluded from the final analysis only for prespecified, objective criteria, again to reduce conscious or unconscious manipulation of the results. For example, appropriate drop of the laser Doppler flowmetry-measured cerebral blood flow is an essential inclusion criterion for middle cerebral artery occlusion models.
- 4. Randomization: animals should be randomized to treatment groups before any study procedures, including behavioral apparatus habituation or

stroke-induction surgery. This helps prevent conscious or unconscious allocation bias.

- 5. Allocation concealment: Ideally, the investigator performing the stroke does not know the treatment assignment of the subject, to avoid unconscious differences in surgical technique.
- 6. Reporting on excluded animals: Subjects may drop out at any phase of the protocol. Reporting these dropouts helps minimize any manipulation of the final results.
- 7. Blinded assessment of outcome: Investigators must remain unaware of group assignments when measuring any outcome. This is especially true of subjective assessments, such as behavior scoring, but applies also to semi-automated morphometry.
- 8. Age: very few patients in stroke trials are younger than 20 years old. Almost all preclinical stroke modeling is performed in rodents 2 to 3 months old, which corresponds to a human age of 15 to 20 years.⁴⁸ At some point before launching human clinical trials, preclinical testing of therapies should include studies in rodents at least 10 months old (corresponding to middle-aged humans).
- 9. Sex: Molecules showing promise in male rodents should also be evaluated in aged female or oopho-rectimized female rodents to demonstrate efficacy in both sexes.
- 10. Comorbidities: Almost two-thirds of patients with stroke in trials are hypertensive. Almost all preclinical testing of therapies are performed in normotensive rodents. Investigators should consider the added value of preclinical testing in hypertensive rodents. Hyperglycemia due to diabetes mellitus is another comorbidity known to influence stroke outcome.
- Multiple laboratories: Before initiating clinical trials, it may be reasonable to test candidate cerebroprotectants for efficacy in multiple laboratories, and in multiple species.^{39,43}
- 12. Gyrencephalic species: Candidate stroke treatments have rarely been tested or shown to be effective in gyrencephalic species such as nonhuman primate stroke models.^{8,49} It remains to be proven whether such demonstration of efficacy in primates will prove superior or complementary to rodent models in predicting success for the candidate therapy in clinical trials.
- 13. Circadian context: Circadian biology affects all aspects of mammalian physiology.⁵⁰ Almost 80% of all currently approved drugs hit targets that show circadian rhythm.⁵¹ It is now well accepted that circadian biology profoundly affects cardio-vascular medicine and immunology.^{52,53} Hence, it may be reasonable to consider the effects of circadian rhythms on stroke therapeutics. Over 90%

of patients with stroke in trials are enrolled in the daytime. For diurnal humans, this is the active (awake) phase. Almost all preclinical testing of therapies is also performed in rodents during the daytime, but for nocturnal rodents, this is their inactive (sleep) phase. A recent study found that stroke evolution and neuroprotectant effects may be different during active versus inactive phases in rodent stroke models.⁵⁴

In conclusion, workshop no. 1 of STAIR XI considered and presented a new paradigm for the evaluation of putative therapies that may work together with recanalization to improve outcome after stroke. This paradigm is presented for the consideration of the larger stroke research community and for further testing and validation.

ARTICLE INFORMATION

Affiliations

Department of Physiology and Neuroscience, Department of Neurology, Keck School of Medicine, Los Angeles (P.L.). Radcliffe Department of Medicine, University of Oxford, Oxford (A.B.). School of Life Sciences, University of Warwick, Coventry (J.B.,). Beth Israel Deaconess Medical Center, Harvard Medical School, Boston (M.F.).

Sources of Funding

Dr Lyden is supported by the NINDS grants U24 NS113452and R01NS075930.

Disclosures

Dr Lyden is the Principal Investigator of the National Institute of Health (NIH) sponsored Stroke Preclinical Assessment Network; a Data Safety Monitoring Board member for the Basilar Artery International Cooperation Study (unpaid) and the PORTICO (PORTICO Re-sheathable Transcatheter Aortic Valve System US IDE Trial; Baim Institute); received royalties for Thrombolytic Therapy for Acute Stroke, 3rd Ed, Springer Press; and serves as a consultant to various plaintiff and defense legal firms and Apex Innovations. Dr Buchan reports other from Brainomix during the conduct of the study. Dr Fisher reports personal fees from ALLM Inc outside the submitted work. The other author reports no conflicts.

Supplemental Materials

List of Contributors

Downloaded from http://ahajournals.org by on August 3, 202

REFERENCES

- Moskowitz MA, Lo EH, ladecola C. The science of stroke: mechanisms in search of treatments. *Neuron*. 2010;67:181–198. doi: 10.1016/j. neuron.2010.07.002
- Asahi M, Wang X, Mori T, Sumii T, Jung JC, Moskowitz MA, Fini ME, Lo EH. Effects of matrix metalloproteinase-9 gene knock-out on the proteolysis of blood-brain barrier and white matter components after cerebral ischemia. J Neurosci. 2001;21:7724–7732.
- Mullen MT, Pisapia JM, Tilwa S, Messé SR, Stein SC. Systematic review of outcome after ischemic stroke due to anterior circulation occlusion treated with intravenous, intra-arterial, or combined intravenous+intra-arterial thrombolysis. *Stroke*. 2012;43:2350–2355. doi: 10.1161/STROKEAHA.111. 639211
- Endres M, Engelhardt B, Koistinaho J, Lindvall O, Meairs S, Mohr JP, Planas A, Rothwell N, Schwaninger M, Schwab ME, et al. Improving outcome after stroke: overcoming the translational roadblock. *Cerebrovasc Dis.* 2008;25:268–278. doi: 10.1159/000118039
- Xiong Y, Mahmood A, Chopp M. Angiogenesis, neurogenesis and brain recovery of function following injury. *Curr Opin Investig Drugs*. 2010;11:298–308.
- Lapchak PA. Emerging therapies: pleiotropic multi-target drugs to treat stroke victims. *Transl Stroke Res.* 2011;2:129–135. doi: 10.1007/ s12975-011-0074-4

- Griffin JH, Mosnier LO, Fernández JA, Zlokovic BV. 2016 Scientific sessions sol sherry distinguished lecturer in thrombosis: thrombotic stroke: neuroprotective therapy by recombinant-activated protein C. Arterioscler Thromb Vasc Biol. 2016;36:2143–2151. doi: 10.1161/ATVBAHA.116.308038
- Cook DJ, Teves L, Tymianski M. Treatment of stroke with a PSD-95 inhibitor in the gyrencephalic primate brain. *Nature*. 2012;483:213–217. doi: 10.1038/nature10841
- Cui H, Hayashi A, Sun H-S, Belmares MP, Cobey C, Phan T, Schweizer J, Salter MW, Wang YT, Tasker RA, et al. Pdz protein interactions underlying nmda receptor-mediated excitotoxicity and neuroprotection by psd-95 inhibitors. *J Neuroscience*. 2007;27:9901–9915.
- 10. Lyden P, Pryor KE, Coffey CS, Cudkowicz M, Conwit R, Jadhav A, Sawyer RN, Jr., Claassen J, Adeoye O, Song S, et al. Final results of the rhapsody trial: A multi-center, phase 2 trial using a continual reassessment method to determine the safety and tolerability of 3k3a-apc, a recombinant variant of human activated protein c, in combination with tissue plasminogen activator, mechanical thrombectomy or both in moderate to severe acute ischemic stroke. Ann Neurol. 2019;85:125–136. doi: 10.1002/ana.25383
- Hill MD, Goyal M, Menon BK, Nogueira RG, McTaggart RA, Demchuk AM, Poppe AY, Buck BH, Field TS, Dowlatshahi D, et al; ESCAPE-NA1 Investigators. Efficacy and safety of nerinetide for the treatment of acute ischaemic stroke (ESCAPE-NA1): a multicentre, double-blind, randomised controlled trial. *Lancet.* 2020;395:878–887. doi: 10.1016/S0140-6736(20)30258-0
- Lyden PD, Pryor KE, Minigh J, Davis TP, Griffin JH, Levy H, Zlokovic BV. Stroke treatment With PAR-1 agents to decrease hemorrhagic transformation. *Front Neurol.* 2021;12:593582. doi: 10.3389/fneur.2021.593582
- Ames A, Wright RL, Kowada M, Thurston JM, Majno G. Cerebral ischemia 2. No-reflow phenomenon. Am J Path. 1968;52:437.
- Lyden PD, Lamb J, Kothari S, Toossi S, Boitano P, Rajput PS. Differential effects of hypothermia on neurovascular unit determine protective or toxic results: toward optimized therapeutic hypothermia. J Cereb Blood Flow Metab. 2018;39:1693–1709. doi: 10.1177/0271678X18814614
- Mordecai Y, Globus T, Busto R. Martinez E, Valdes I, Ginsberg MD. Excitotoxic index - a biochemical marker of selective vulnerability. *Stroke*. 1991;22:128.
- Collins RC, Dobkin BH, Choi DW. Selective vulnerability of the brain: new insights into the pathophysiology of stroke. *Ann Intern Med.* 1989;110:992– 1000. doi: 10.7326/0003-4819-110-12-992
- Kirino T, Sano K. Selective vulnerability in the gerbil hippocampus following transient ischemia. Acta Neuropathol. 1984;62:201–208. doi: 10.1007/BF00691853
- Mabuchi T, Lucero J, Feng A, Koziol JA, del Zoppo GJ. Focal cerebral ischemia preferentially affects neurons distant from their neighboring microvessels. J Cereb Blood Flow Metab. 2005;25:257–266. doi: 10.1038/sj.jcbfm.9600027
- Xing C, Lo EH. Help-me signaling: Non-cell autonomous mechanisms of neuroprotection and neurorecovery. *Prog Neurobiol.* 2017;152:181–199. doi: 10.1016/j.pneurobio.2016.04.004
- Rajput PS, Lamb J, Kothari S, Pereira B, Soetkamp D, Wang Y, Tang J, Van Eyk JE, Mullins ES, Lyden PD. Neuron-generated thrombin induces a protective astrocyte response via protease activated receptors. *Glia.* 2020;68:246–262. doi: 10.1002/glia.23714
- Lu Y, Sareddy GR, Wang J, Zhang O, Tang FL, Pratap UP, Tekmal RR, Vadlamudi RK, Brann DW. Neuron-derived estrogen is critical for astrocyte activation and neuroprotection of the ischemic brain. *J Neurosci.* 2020;40:7355–7374. doi: 10.1523/JNEUROSCI.0115-20.2020
- Wang J, Sareddy GR, Lu Y, Pratap UP, Tang F, Greene KM, Meyre PL, Tekmal RR, Vadlamudi RK, Brann DW. Astrocyte-derived estrogen regulates reactive astrogliosis and is neuroprotective following ischemic brain injury. *J Neuroscience*. 2020;40:9751–9771. doi: 10.1523/ JNEUROSCI.0888-20.2020
- Pitt J, Wilcox KC, Tortelli V, Diniz LP, Oliveira MS, Dobbins C, Yu XW, Nandamuri S, Gomes FCA, DiNunno N, et al. Neuroprotective astrocytederived insulin/insulin-like growth factor 1 stimulates endocytic processing and extracellular release of neuron-bound Aβ oligomers. *Mol Biol Cell*. 2017;28:2623–2636. doi: 10.1091/mbc.E17-06-0416
- Hayakawa K, Esposito E, Wang X, Terasaki Y, Liu Y, Xing C, Ji X, Lo EH. Transfer of mitochondria from astrocytes to neurons after stroke. *Nature*. 2016;535:551–555. doi: 10.1038/nature18928
- Jha MK, Seo M, Kim JH, Kim BG, Cho JY, Suk K. The secretome signature of reactive glial cells and its pathological implications. *Biochim Biophys Acta*. 2013;1834:2418–2428. doi: 10.1016/j.bbapap.2012.12.006
- Barreto G, White RE, Ouyang Y, Xu L, Giffard RG. Astrocytes: targets for neuroprotection in stroke. *Cent Nerv Syst Agents Med Chem.* 2011;11:164– 173. doi: 10.2174/187152411796011303

SPECIAL REPORT

- Wang Y, Luo W, Reiser G. Activation of protease-activated receptors in astrocytes evokes a novel neuroprotective pathway through release of chemokines of the growth-regulated oncogene/cytokine-induced neutrophil chemoattractant family. *Eur J Neurosci.* 2007;26:3159–3168. doi: 10.1111/j.1460-9568.2007.05938.x
- Lin CH, Cheng FC, Lu YZ, Chu LF, Wang CH, Hsueh CM. Protection of ischemic brain cells is dependent on astrocyte-derived growth factors and their receptors. *Exp Neurol.* 2006;201:225–233. doi: 10.1016/j. expneurol.2006.04.014
- Trendelenburg G, Dirnagl U. Neuroprotective role of astrocytes in cerebral ischemia: focus on ischemic preconditioning. *Glia*. 2005;50:307–320. doi: 10.1002/glia.20204
- Savitz SI, Baron JC, Fisher M; STAIR X Consortium. Stroke treatment academic industry roundtable X: Brain cytoprotection therapies in the reperfusion era. Stroke. 2019;50:1026–1031. doi: 10.1161/STROKEAHA.118.023927
- National Institute of Neurological Disorders and Stroke rt-PA Stroke Study Group. Tissue plasminogen activator for acute ischemic stroke. N Engl J Med. 1995;333:1581–1587. doi: 10.1056/NEJM199512143332401
- Hacke W, Kaste M, Fieschi C, von Kummer R, Davalos A, Meier D, Larrue V, Bluhmki E, Davis S, Donnan G, et al. Randomised double-blind placebo-controlled trial of thrombolytic therapy with intravenous alteplase in acute ischaemic stroke (ECASS II). Second European-Australasian Acute Stroke Study Investigators. *Lancet.* 1998;352:1245–1251. doi: 10.1016/s0140-6736(98)08020-9
- 33. Lansberg MG, Lee J, Christensen S, Straka M, De Silva DA, Mlynash M, Campbell BC, Bammer R, Olivot JM, Desmond P, et al. RAPID automated patient selection for reperfusion therapy: a pooled analysis of the Echoplanar Imaging Thrombolytic Evaluation Trial (EPITHET) and the Diffusion and Perfusion Imaging Evaluation for Understanding Stroke Evolution (DEFUSE) Study. *Stroke.* 2011;42:1608–1614. doi: 10.1161/STROKEAHA.110.609008
- Thomalla G, Simonsen CZ, Boutitie F, Andersen G, Berthezene Y, Cheng B, Cheripelli B, Cho TH, Fazekas F, Fiehler J, et al; WAKE-UP Investigators. MRI-guided thrombolysis for stroke with unknown time of onset. N Engl J Med. 2018;379:611–622. doi: 10.1056/NEJMoa1804355
- Albers GW, Marks MP, Kemp S, Christensen S, Tsai JP, Ortega-Gutierrez S, McTaggart RA, Torbey MT, Kim-Tenser M, Leslie-Mazwi T, et al; DEFUSE 3 Investigators. Thrombectomy for stroke at 6 to 16 hours with selection by perfusion imaging. *N Engl J Med.* 2018;378:708–718. doi: 10.1056/ NEJMoa1713973
- Ma H, Campbell BCV, Parsons MW, Churilov L, Levi CR, Hsu C, Kleinig TJ, Wijeratne T, Curtze S, Dewey HM, et al; EXTEND Investigators. Thrombolysis guided by perfusion imaging up to 9 hours after onset of stroke. N Engl J Med. 2019;380:1795–1803. doi: 10.1056/NEJMoa1813046
- Nogueira RG, Jadhav AP, Haussen DC, Bonafe A, Budzik RF, Bhuva P, Yavagal DR, Ribo M, Cognard C, Hanel RA, et al; DAWN Trial Investigators. Thrombectomy 6 to 24 hours after stroke with a mismatch between deficit and infarct. *N Engl J Med.* 2018;378:11–21. doi: 10.1056/NEJMoa1706442
- Sena E, van der Worp HB, Howells D, Macleod M. How can we improve the pre-clinical development of drugs for stroke? *Trends Neurosci.* 2007;30:433–439. doi: 10.1016/j.tins.2007.06.009

- Macleod MR, Fisher M, O'Collins V, Sena ES, Dirnagl U, Bath PM, Buchan A, van der Worp HB, Traystman R, Minematsu K, et al. Good laboratory practice: preventing introduction of bias at the bench. *Stroke*. 2009;40:e50– e52. doi: 10.1161/STROKEAHA.108.525386
- Lapchak PA, Zhang JH, Noble-Haeusslein LJ. RIGOR guidelines: escalating STAIR and STEPS for effective translational research. *Transl Stroke Res.* 2013;4:279–285. doi: 10.1007/s12975-012-0209-2
- Bosetti F, Koenig JI, Ayata C, Back SA, Becker K, Broderick JP, Carmichael ST, Cho S, Cipolla MJ, Corbett D, et al. Translational stroke research: vision and opportunities. *Stroke.* 2017;48:2632–2637. doi: 10.1161/ STROKEAHA.117.017112
- Stroke Therapy Academic Industry Roundtable (STAIR). Recommendations for standards regarding preclinical neuroprotective and restorative drug development. *Stroke*. 1999;30:2752. doi: 10.1161/01.str.30.12.2752
- Fisher M, Feuerstein G, Howells DW, Hurn PD, Kent TA, Savitz SI, Lo EH; STAIR Group. Update of the stroke therapy academic industry roundtable preclinical recommendations. *Stroke*. 2009;40:2244–2250. doi: 10.1161/STROKEAHA.108.541128
- van der Worp HB, Howells DW, Sena ES, Porritt MJ, Rewell S, O'Collins V, Macleod MR. Can animal models of disease reliably inform human studies? *PLoS Med.* 2010;7:e1000245. doi: 10.1371/journal.pmed.1000245
- Sena ES, van der Worp HB, Bath PM, Howells DW, Macleod MR. Publication bias in reports of animal stroke studies leads to major overstatement of efficacy. *PLoS Biol.* 2010;8:e1000344. doi: 10.1371/journal.pbio.1000344
- Voelkl B, Vogt L, Sena ES, Würbel H. Reproducibility of preclinical animal research improves with heterogeneity of study samples. *PLoS Biol.* 2018;16:e2003693. doi: 10.1371/journal.pbio.2003693
- Kilkenny C, Browne WJ, Cuthill IC, Emerson M, Altman DG. Improving bioscience research reporting: the ARRIVE guidelines for reporting animal research. *PLoS Biol.* 2010;8:e1000412. doi: 10.1371/ journal.pbio.1000412
- Sengupta P. The laboratory rat: relating its age with human's. Int J Prev Med. 2013;4:624–630.
- Marshall Jonathan WB, Cummings Rosalyn M, Bowes Laura J, Ridley Rosalind M, Green AR. Functional and histological evidence for the protective effect of nxy-059 in a primate model of stroke when given 4 hours after occlusion. *Stroke*. 2003;34:2228.
- Cederroth CR, Albrecht U, Bass J, Brown SA, Dyhrfjeld-Johnsen J, Gachon F, Green CB, Hastings MH, Helfrich-Förster C, Hogenesch JB, et al. Medicine in the fourth dimension. *Cell Metab.* 2019;30:238–250. doi: 10.1016/j.cmet.2019.06.019
- Rijo-Ferreira F, Takahashi JS. Genomics of circadian rhythms in health and disease. *Genome Med.* 2019;11:82. doi: 10.1186/s13073-019-0704-0
- Paschos GK, FitzGerald GA. Circadian clocks and vascular function. *Circ* Res. 2010;106:833–841. doi: 10.1161/CIRCRESAHA.109.211706
- Scheiermann C, Kunisaki Y, Frenette PS. Circadian control of the immune system. Nat Rev Immunol. 2013;13:190–198. doi: 10.1038/nri3386
- Esposito E, Li W, T Mandeville E, Park JH, Şencan I, Guo S, Shi J, Lan J, Lee J, Hayakawa K, et al. Potential circadian effects on translational failure for neuroprotection. *Nature*. 2020;582:395–398. doi: 10.1038/ s41586-020-2348-z